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# PstI

Cat # □ RE109S □ RE109L Pack Size: □ 500U □ 1000U Volume: □ 50 µL □ 100 µL Storage: -20°C **Recognition Sequence:** 5' CTGCAG 3' 3' GACGTC 5'



## Kit content:

S.No	Component	RE109S	RE109L
1	PstI	500U/50µL(10U/µL)	1000U/100µL(10U/µL)
2	10X Universal Buffer	500 μL	1 mL

## Introduction

PstI is a type 2 restriction enzyme that recognizes and cuts the DNA sequence 5'-CTGCA^G-3'. It is commonly used in molecular biology for the digestion of DNA fragments during cloning experiments. It is sourced from an *E. coli* strain, that carries the cloned PstI gene from *Providencia stuartii*.

#### **Features**

- Assayed on λ DNA.
- Ligation/recutting assay: After 20-fold overdigestion with enzyme, >90% of the DNA fragments can be ligated and recut.
- Overdigestion assay: No nonspecific activity detected after incubation of 1  $\mu$ g of  $\lambda$  DNA with 20 units of PstI for 16 hours.

## Protocol:

- The enzyme should not exceed 10 % of total reaction volume.
- Add enzyme as last component. Mix components well before adding enzyme. After enzyme addition, mix gently by pipetting. Do not vortex
- Incubate 60 min. at 37 °C.

S.No	Component	50 µl Reaction
1	DNA	1 µg
2	10X Universal Buffer	5 µL
3	PstI	2-5 units
4	Nuclease-free Water	to 50µL

## Enzyme Inactivation:

Stop reaction by alternatively:

- Addition of 2.1 µl EDTA pH 8.0 [0.5 M], final 20 mM
- Heat Inactivation (20 min. at 65 °C)
- Spin Column DNA Purification (e.g. Cat.-No. <u>PUR13-50</u>)
- Gel Electrophoresis and Single Band Excision (e.g. Cat.-No. <u>PUR12-50</u>)
- Phenol-Chloroform Extraction or Ethanol Precipitation.

Supplied in :10mM Tris-Hcl (pH 7.6), 50mM NaCl, 0.1mM EDTA, 1mM DTT, 100µg/ml, BSA, 50% Glycerol.

Unit definition :One unit is defined as the amount of restriction enzyme required to completely digest 1 µg of lambda DNA in 1 hour at 37°C in a total reaction volume of 50 µl.

## Quality Control Assays:

- Ligation of DNA fragments: DNA fragments are produced by an excessive over digestion of substrate DNA with each restriction endonuclease. These fragments are then ligated with T4 DNA Ligase at a 5' termini concentration of 0.1-1.0 µM. The ligated fragments are then recut with the same restriction endonuclease. Ligation can only occur if the 3' and 5' termini are left intact, and only those molecules with a perfectly restored recognition site can be recleaved. A normal banding pattern after cleavage indicates that both the 3' and 5' termini are intact, and the enzyme preparation is free of detectable exonucleases and phosphatases.
- DNA digestion with restriction enzymes may be affected by some types of methylation.
- In general, it is recommended to use 5–10 units of enzyme per µg of plasmid DNA, and 10–20 units for genomic DNA in a 1-hour digest. Enzyme volume should not exceed 10% of the total reaction volume to prevent star activity.
- All preparations are assayed for contaminating endonuclease, 3'-exonuclease, 5' exonuclease/ 5' phosphatase, as well as nonspecific single- and doublestranded DNase activities.